



The identification of structurally novel, selective, orally bioavailable positive modulators of mGluR2

Pier L. D'Alessandro^a, Corrado Corti^b, Adelheid Roth^b, Annarosa Ugolini^b, Anna Sava^b, Dino Montanari^b, Federica Bianchi^b, Stephen L. Garland^c, Ben Powney^c, Emma L. Koppe^c, Magalie Rocheville^c, Greg Osborne^c, Paloma Perez^d, Jesús de la Fuente^d, Maite De Los Frailes^d, Paul W. Smith^a, Clive Branch^a, David Nash^a, Stephen P. Watson^{a,*}

^a GlaxoSmithKline, Neurosciences Centre of Excellence for Drug Discovery, New Frontiers Science Park, Harlow, UK

^b GlaxoSmithKline, Neurosciences Centre of Excellence for Drug Discovery, Medicines Research Centre, Verona, Italy

^c GlaxoSmithKline, Molecular Discovery Research, New Frontiers Science Park, Harlow, UK

^d GlaxoSmithKline, Molecular Discovery Research, Tres Cantos, Madrid, Spain

ARTICLE INFO

Article history:

Received 6 October 2009

Revised 6 November 2009

Accepted 9 November 2009

Available online 26 November 2009

Keywords:

Metabotropic glutamate receptor ligand

Metabotropic glutamate receptor

Positive allosteric modulator

mGluR

mGlu

mGluR2

ABSTRACT

The optimisation of an HTS hit series (**1**) leading to the identification of structurally novel, selective, orally bioavailable mGluR2 positive modulators GSK1331258 and GSK1331268 is described. Structure–activity relationships, attenuation of dopaminergic activity, and potentiation of mGluR2 responses in rat hippocampal MPP-DG synapses are also reported.

© 2009 Elsevier Ltd. All rights reserved.

There is a strong body of evidence that modulation of metabotropic glutamate receptors could afford novel approaches for the treatment of neurodegenerative and neuropsychiatric diseases.^{1–4} The type 2 metabotropic glutamate receptor is of particular interest, indeed, a prodrug of the orthosteric mGluR2/3 agonist LY404039 (Lilly) exhibited statistically significant improvements in both positive and negative symptoms of schizophrenia in phase II trials.⁵ Additionally a study using receptor deficient mice showed that antipsychotic effects of the mGluR2/3 receptor agonist LY404039 are dependent on mGluR2 and not mGluR3 receptors.⁶

Positive allosteric modulation could offer a superior mechanism of potentiating mGluR2 function than classical orthosteric agonism. The advantages positive modulation may afford, in terms of enhanced selectivity and spatial and temporal control, have been described extensively elsewhere^{7,8} suffice it to say that this mechanism has received a great deal of attention in recent patent and scientific literature. Notable recent reports of mGluR2 positive modulators include the pioneering work of Lilly, exemplified by pyridylsulfonamides LY487379^{10,9–13} and, a promising report¹⁴ of

compounds such as **2** from Pfizer (Fig. 2), which prompted us to report the work from our laboratories.

Herein we describe a novel series of 2-piperidine/piperazine-methylene-benzimidazole mGluR2 positive modulators derived from a high throughput screening (HTS) hit series exemplified by compound **1** (Fig. 1). The synthesis, structure–activity relationships (SAR) investigation, and the control of dopaminergic off-target activities of this series is described: culminating in the identification of the selective, orally bioavailable mGluR2 positive modulators GSK1331258 and GSK1331268.

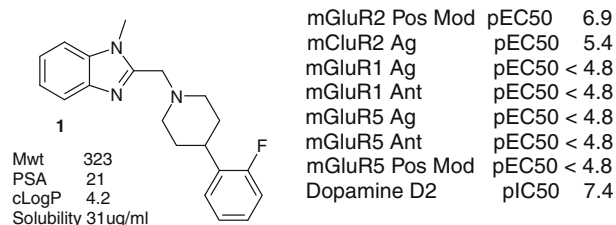


Figure 1. Structure and profile of HTS hit compound **1**.

* Corresponding author. Tel.: +44 1279 875004.

E-mail address: steve.p.watson@gsk.com (S.P. Watson).

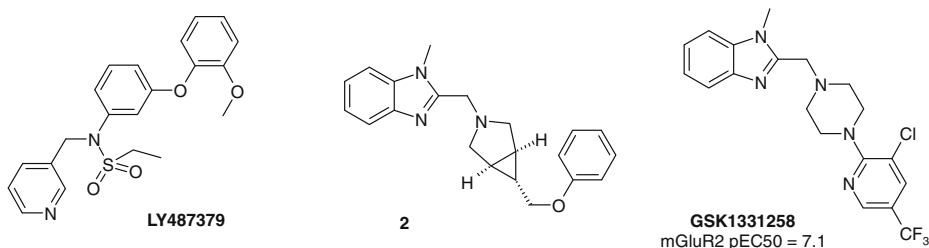
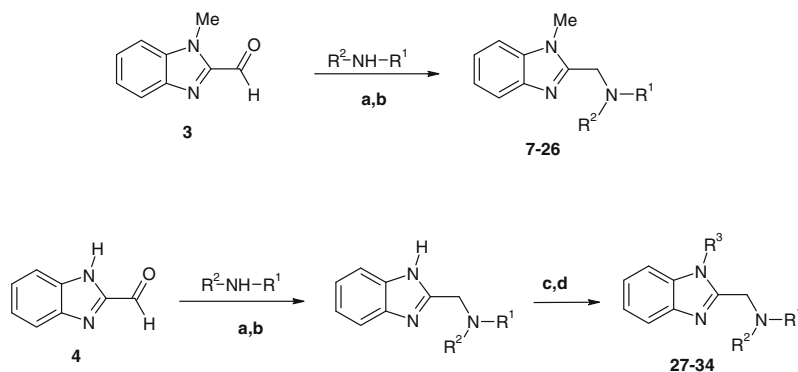


Figure 2. Exemplar mGluR2 positive modulators and GSK1331258.



Scheme 1. Reagents and conditions: (a) DCM/AcOH = 98:2 (V/V), rt, 15 min; (b) MP-BH(OAc)₃ (5.0 equiv), microwave irradiation, 110 °C, 10 min; 50–90%; (c) 60% NaH in mineral oil, DMF/THF = 30:70 (V/V), 10 min at rt; (d) R³X, microwave irradiation, 100 °C, 10 min (X = Cl, Br), 40–75%.

From a HTS in a chimeric CHO cell line using a FLIPR readout¹⁵ several promising mGluR2 modulator chemotypes were identified. Compound **1** was selected for further progression based on its displaying the best balance of pharmacology, selectivity, developability and properties suitable to afford good CNS penetration (Mol. wt <400, *c* Log *P* <4.5 and PSA <80). It was anticipated that dopaminergic activity associated with this template could be readily attenuated.

The desired mechanism of action of this compound/chemotype was confirmed by blockade of the positive modulation by the orthosteric mGluR2 antagonist¹⁶ LY341495. A medicinal chemistry programme was initiated with the aim of identifying selective, orally bioavailable and brain penetrant mGluR2 positive modulators suitable for progression to studies in *in vivo* models.

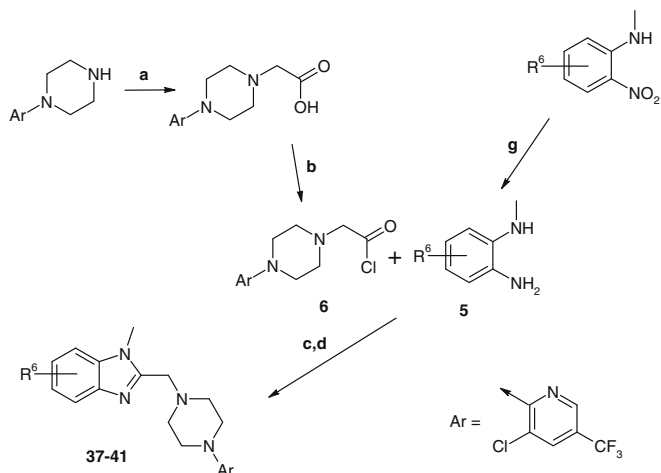
Compounds were typically prepared according to the chemistry shown in Schemes 1 and 2.¹⁷

Simple reductive amination of 1-methyl-3-formyl benzimidazole (**3**) with appropriate amines allowed investigation of the aryl-piperazine/piperidine region of the template (Table 1). Variation of the benzimidazole N-substituent was achieved via a final step alkylation of intermediates generated by reductive amination of 2-formylbenzimidazole (**4**) (Table 2). Variation of the benzimidazole benzosubstituents (Table 2) was effected via reaction of *N*-methyl bis-anilines (**5**) and the corresponding piperazine/piperidine derived glycine (**6**).

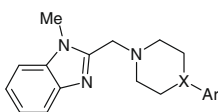
Table 1 illustrates the profiles of a representative set of analogues of lead compound **1**, in which the piperazine, and corresponding piperidine, aryl ring is modified. There are apparent differences in SAR between the piperazines and piperidines. Piperidines are typically more potent mGluR2 positive modulators than the corresponding piperazines (e.g., contrast the potency of the two simple phenyl analogues **7** and **17**) but in some instances potencies are comparable (e.g., 4-trifluoromethyl analogues **11** and **22**). Most compounds also demonstrated weak agonism of the receptor, this could be due to low level allosteric agonism (but effects due to contaminating traces of glutamate in the assay

preparation cannot be discounted). Compounds typically potentiated an EC₂₀ of glutamate to 70–100% of the glutamate EC₁₀₀ response, but there is no apparent correlation between this efficacy of potentiation and pEC₅₀.

Close analogues of the chemotype exemplified by compound **1** are known to display activities at dopaminergic receptors,^{18–20} hence it was important to investigate and attenuate any dopaminergic activity²¹ such that dopaminergic pharmacology could not confound readout in any downstream *in vivo* models. Data in Table 1 illustrates that piperidine based analogues of compound **1** indeed typically demonstrated significant D₂ and D₃ antagonism. However this activity could be successfully attenuated in the



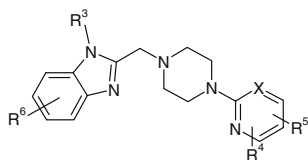
Scheme 2. Reagents and conditions: (a) BrCH₂COOH, MeCN, rt, overnight, 62%; (b) 1-chloro-*N,N*,2-trimethyl-1-propen-1-amine MeCN, rt, 40 min; (c) MeCN, 20 min at rt; (d) MeCN, microwave irradiation, 140 °C, 20 min; (b–d in one pot-11–25% overall); (g) Fe, NH₄Cl, 50% MeOH/H₂O (V/V), 2 h at reflux, 40–60%.

Table 1Positive modulation of EC₂₀ glutamate¹⁵ at mGluR2 and dopaminergic antagonism²⁰ by piperazine and piperidine analogues

Compound	Ar	X	mGluR2 pos. mod. pEC ₅₀	mGluR2 pos. mod. max.	mGluR2 agonist. pEC ₅₀	D2 pK _i	D3 pK _i	D4 pK _i
7	Phenyl	CH	6.7	89	5.2	7.6	6.9	7.0
1	2-F-Phenyl	CH	6.9	91	5.4	7.4	7.0	
8	4-Chloro-phenyl	CH	7.1	91	5.3	7.8	6.5	7.2
9	2-Trifluoromethyl-phenyl	CH	6.5	94	5.2	<5.5	6.3	
10	3-Trifluoromethyl-phenyl	CH	7	94	5.6	6.9	<5	6.6
11	4-Trifluoromethyl-phenyl	CH	7	110	5.7	6.6	<5	6.9
12	3-Chloro-phenyl	CH	7	92	5.4	7.4	5.8	6.6
13	1,3-Benzothiazole-2-yl	CH	6.8	76	5.5	6.4	<5	8.1
14	1,2-Benzisothiazole-3-yl	CH	6.7	84	5.6	7.1	5.4	6.7
15	3-Chloro-5-(trifluoromethyl)-2-pyridinyl	CH	7.4	70	6.2	6.1	5.8	5.6
16	2-Pyridyl	CH	<5.5	n/a	<4.8			
17	Phenyl	N	5.3	100	<4.8			
18	2-F-Phenyl	N	6.1	64	5.2			
19	4-Chloro-phenyl	N	6.1		<4.8	<5	<5	6.9
20	2-Trifluoromethyl-phenyl	N	6.5	84	<4.8			6.2
GSK1331258	3-Chloro-5-(trifluoromethyl)-2-pyridinyl	N	7.1	71	5.7	5.3	5.3	6.2
GSK1331268	5-Chloro-2-pyrimidinyl	N	6.9	69	5.9	<5	<5	5.6
21	4-Chloro-2-fluoro-phenyl	N	6.9	69	5.6			6.6
22	4-Trifluoromethyl-phenyl	N	6.7	84	5.2	<5	<5	6.4
23	1,3-Benzothiazole-2-yl	N	6.5	88	5.7	<5	<5	6.1
24	5-(Trifluoromethyl)-2-pyridinyl	N	6.3	81	5.2	<5	<5	6.0
25	2-Pyridyl	N	4.8	100				
26	4-Cyanophenyl	N	<4.9	n/a				

majority of the piperazines. D₄ antagonism was more persistent within the analogues, but was reduced to a very low level in compounds such as GSK1331268.

Based on the data in Table 1, the 5-chloro-2-pyrimidinyl and 3-chloro-5-(trifluoromethyl)-2-pyridinyl substituted piperazine fragments of compounds GSK1331258 and GSK1331268 were selected for use in further studies (see Table 2), as they represented the best balance of high mGluR2 activity and low dopaminergic activity.

Table 2FLIPR pEC₅₀ positive modulator activity as potentiation of EC₂₀ glutamate¹⁵

	X	R ³	R ⁴	R ⁵	R ⁶	mGluR2 pos. mod. pEC ₅₀
27	C	–Et	2-Cl	5-CF ₃	H	7.2
28	C	–CH ₂ Ph	2-Cl	5-CF ₃	H	6.9
29	N	–CH ₂ Ph	5-Cl	H	H	6.9
30	C	–(CH ₂) ₂ OCH ₃	2-Cl	5-CF ₃	H	6.8
31	N	–Et	5-Cl	H	H	6.8
32	N	–(CH ₂) ₂ OCH ₃	5-Cl	H	H	6.3
33	N	–nProp	5-Cl	H	H	6.1
34	C	–nProp	2-Cl	5-CF ₃	H	5.6
35	C	–H	2-Cl	5-CF ₃	H	4.9
36	N	–H	5-Cl	H	H	4.9
37	C	Me	2-Cl	5-CF ₃	5-OMe	6.6
38	C	Me	2-Cl	5-CF ₃	5-CN	7.3
39	C	Me	2-Cl	5-CF ₃	5-CF ₃	5.7
40	C	Me	2-Cl	5-CF ₃	6-F	6.9
41	C	Me	2-Cl	5-CF ₃	5-F	6.7

Compounds **27–34** (Table 2) illustrate that increasing the size of the benzimidazole N-substituent affords little enhancement of mGluR2 pharmacology. Interestingly the two N-unsubstituted analogues **35** and **36** have effectively no mGluR2 activity.

Table 2 also reports data for modifications to benzimidazole ring substitution of 3-chloro-5-(trifluoromethyl)-2-pyridinyl piperazines (**37–41**). Similarly to further Benzimidazole N-substitution, these ring substitutions conferred no significant improvement on mGluR2 pharmacology.

In contrast to the majority of analogues GSK1331258 and GSK1331268 demonstrated good in vitro metabolic stability (data not shown) and were selected for progression for in vivo pharmacokinetic investigation. In rat, in a 1 mg/kg oral cassette study both compounds showed a promising profile, viz. GSK1331258 exhibited low clearance (CL_b 18.0 ml/min/kg), a half life of 4.1 h, a brain:blood ratio of 3.2 with brain C_{max} of 147 ng/g, and bioavailability of 58%; GSK1331268 exhibited moderate clearance (CL_b 32.0 ml/min/kg), a half life of 1.0 h, a brain–blood ratio of 1.3 with brain C_{max} of 32 ng/g and bioavailability of 22%.

Furthermore GSK1331258 and GSK1331268 preserved the mGluR selectivity observed in initial hit compound **1**, with neither compound displaying significant activity at mGluRs1, 4 or 5 in agonist, antagonist or positive modulator modes (data not shown).

The pharmacology of GSK1331268 was characterised further on Medial Perforant-Path Dentate Gyrus (MPP-DG) Synapses in rat hippocampal slices,²² in which synaptic transmission is inhibited by activation of group II mGluR receptors.²³

Orthosteric agonist LY354740^{24,25} inhibits MPP-DG fEPSP with an EC₅₀ of 134 nM. Co-application of LY354740 with GSK1331268 (10 μM) shifts the concentration–response curve for LY354740 to the left by threefold, to 43 nM (see Fig. 3).

In summary a successful SAR investigation of a HTS hit series (**1**) was conducted. Understanding of factors controlling mGluR2 activity and dopaminergic activity allowed the identification of structurally novel, selective,²⁶ and orally bioavailable mGluR2 positive

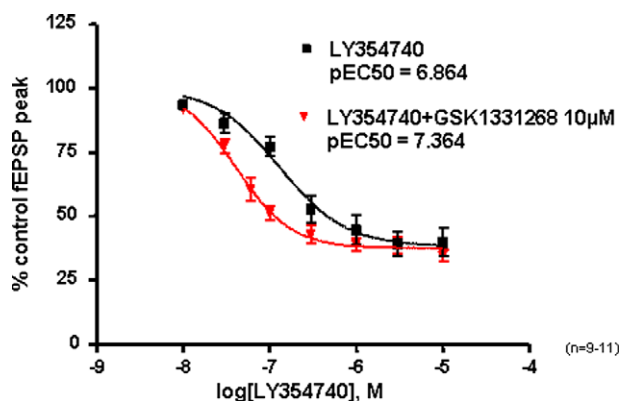


Figure 3. The effect of GSK1331268 on excitatory post-synaptic potentials (fEPSP) at MPP-DG synapses in rat hippocampal slices (data points indicate the mean $\pm$ SEM of fEPSP peak amplitude expressed as a percentage of predrug control baseline; $n = 9$ –11 slices per data point).

modulators GSK1331258 and GSK1331268. Indeed these compounds have suitable profiles for investigation of the effects mGluR2 positive modulation in in vivo disease models.

References and notes

- Lam, A. G.; Soriano, M. A.; Monn, J. A.; Schoepp, D. D.; Lodge, D.; McCulloch, J. *Neurosci. Lett.* **1998**, *2540*, 121.
- Kingston, A. E.; O'Neill, M. J.; Lam, A.; Bales, K. R.; Monn, J. A.; Schoepp, D. D. *Eur. J. Pharmacol.* **1999**, *377*, 155.
- Helton, D. R.; Tizzano, J. P.; Monn, J. A.; Schoepp, D. D.; Kallman, M. J. *J. Pharmacol. Exp. Ther.* **1998**, *284*, 651.
- Chavez-Noriega, L. E.; Schaffhauser, H.; Campbell, U. C. *Curr. Drug Targets CNS Neurol. Disord.* **2002**, *1*, 261.
- Patil, S. T.; Zhang, L.; Martenyi, F.; Lowe, S. L.; Jackson, K. A.; Andreev, B. V.; Avedisova, A. S.; Bardenstein, L. M.; Gurovich, I. Y.; Morozova, M. A.; Mosolov, S. N.; Neznanov, N. G.; Reznik, A. M.; Smulevich, A. B.; Tochilov, V. A.; Johnson, B. G.; Monn, J. A.; Schoepp, D. D. *Nat. Med.* **2007**, *13*, 1102.
- Fell, M. L.; Svensson, K. A.; Johnson, B. J.; Schoepp, D. D. *J. Pharmacol. Exp. Ther.* **2008**, *326*, 209.
- Kew, J. N. C. *Pharm. Theor.* **2004**, *104*, 233.
- Pin, J. P.; Acher, F. *Curr. Drug Targets CNS. Neurol. Disord.* **2002**, *3*, 297–317.
- Johnson, M. P.; Barda, D.; Britton, T. C.; Emkey, R.; Hornback, W. J.; Jagdmann, G. E.; McKinzie, D. L.; Nisenbaum, E. S.; Tizzano, J. P.; Schoepp, D. D. *Psychopharmacology* **2005**, *179*, 271.
- Coleman, D. S.; Jagdmann, G. E. J.; Johnson, K. W.; Johnson, Michael P.; Large, T. H.; Monn, J. A.; Schoepp, D. D.; Tizzano, J. P.; Barda, D. A.; Britton, T. C.; Dressman, B. A.; Fichtner, M. W.; Henry, S. S.; Hornback, W. J. Patent: WO 2001056990.
- Imogai, H. J.; Cid-Nunez, J. M.; Andres-Gil, J. I.; Trabanco-Suarez, A. A.; Oyarzabal Santamarina, J.; Dautzenberg, F. M.; MacDonald, G. J.; Pullan, S. E.; Luetjens, R. J.; Duvey, G. A. J.; Nhem, V.; F.; Terry, P.; Melikyan, G. 2007; Patent: WO 2007104783.
- Bonnefous, C.; Vernier, J. M.; Hutchinson, J. H.; Gardner, M. F.; Cramer, M.; James, J. K.; Rowe, B. A.; Daggett, L. P.; Schaffhauser, H.; Kamenecka, T. M. J. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 4354.
- Rudd, M. T.; McCauley, J. A. *Curr. Top. Med. Chem.* **2005**, *5*, 869.
- Zhang, L.; Rogers, B. N.; Duplantier, A. J.; McHardy, S. F.; Efremov, I.; Berke, H.; Qian, W.; Zhang, A. Q.; Maklad, N.; Candler, J.; Doran, A. C.; Lazzaro, J. T., Jr.; Ganong, A. H. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 5493.
- mGluR2 pharmacology characterised using a FLIPR (Fluorometric Imaging Plate Reader) assay utilising CHO cells stably expressing both the human mGluR2 receptor and Ga16 G-protein. 11 Point dose–response curves were generated, and for positive modulation in presence of an EC₂₀ of glutamate. Values are typically average of four or more experiments. Positive Modulation Max Response is calculated relative to response of EC₁₀₀ of glutamate.
- Johnson, B. G.; Wright, R. A.; Arnold, M. B.; Wheeler, W. J.; Ornstein, P. L.; Schoepp, D. D. *Neuropharmacology* **1999**, *38*, 1519.
- Structure of final compounds confirmed via H NMR and LC–MS analysis.
- Stewart, A. O.; Cowart, M. D.; Moreland, R. B.; Latshaw, S. P.; Matulenko, M. A.; Bhatia, P. A.; Wang, X.; Daanen, J. F.; Nelson, S. L.; Terranova, M. A.; Namovic, M. T.; Donnelly-Roberts, D. L.; Miller, L. N.; Nakane, M.; Sullivan, J. P.; Brioni, J. D. *J. Med. Chem.* **2004**, *47*, 2348.
- Nakane, M.; Cowart, M. D.; Hsieh, G. C.; Miller, L.; Uchic, M. E.; Chang, R.; Terranova, M. A.; Donnelly-Roberts, D. L.; Namovic, M. T.; Miller, T. R.; Wetter, J. M.; Marsh, K.; Stewart, A. O.; Brioni, J. D.; Moreland, R. B. *Neuropharmacology* **2005**, *49*, 112.
- Cowart, M.; Latshaw, S. P.; Bhatia, P. A.; Daanen, J. F.; Rohde, J. J.; Nelson, S. L.; Patel, M. V.; Kolasa, T.; Nakane, M.; Uchic, M. E.; Miller, L. N.; Terranova, M. A.; Chang, R.; Donnelly-Roberts, D. L.; Namovic, M. T.; Hollingsworth, P. R.; Martino, B. R.; Lynch, J. J.; Sullivan, J. P.; Hsieh, G. C.; Moreland, R. B.; Brioni, J. D.; Stewart, A. O. *J. Med. Chem.* **2004**, *47*, 3853.
- D2 and D3, [35S]-GTPγS assay CHO membranes expressing G0 with hD3 or hD2 transfected; D4 FLIPR assay in HEK293 cells stably expressing hD4 and transiently expressing the chimeric Gqo5 G-protein. Values are typically an average of four or more experiments generated from 11 point dose–response curves.
- Schaffhauser, H.; Rowe, B. A.; Morales, S.; Chavez-Noriega, L. E.; Yin, R.; Jachec, C.; Rao, S. P.; Bain, G.; Pinkerton, A. B.; Vernier, J. M.; Bristow, L. J.; Varney, M. A.; Daggett, L. P. *Mol. Pharmacol.* **2003**, *64*, 798.
- Kew, J. N.; Ducarre, J. M.; Pflimlin, M. C.; Mutel, V.; Kemp, J. A. *Neuropharmacology* **2001**, *40*, 20.
- Schoepp, D. D.; Johnson, B. G.; Wright, R. A.; Salhoff, C. R.; Maynen, G.; Wu, S.; Cocherham, S. L.; Burnett, J. P.; Belegaje, R.; Bleakman, D.; Monn, J. A. *Neuropharmacology* **1997**, *36*, 1.
- Monn, J. A.; Valli, M. J.; Massey, S. M.; Hansen, M. M.; Kress, T. J.; Wepsiec, J. P.; Harkness, A. R.; Grutsch, J. L.; Wright, R. A.; Johnson, B. G.; Andis, S. L.; Kingston, A.; Tomlinson, R.; Lewis, R.; Griffey, K. R.; Tizzano, J. P.; Schoepp, D. D. *J. Med. Chem.* **1999**, *42*, 1027.
- GSK1331258 and GSK1331268 showed no significant activity across a panel of representative serotonergic, histaminergic and adrenergic receptors. Activity at mGluR3 was not determined.